

CHEMICAL CONSTITUENTS OF *Eria spicata*

Liqin Wang,¹ Mingmei Wu,² Jian Huang,¹
Jihua Wang,¹ Yegao Chen,^{1*}
and Benlin Yin¹

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In order to know if *Eria* (Orchidaceae) plants can replace the precious medical *Dendrobium* (Orchidaceae) plants, chemical research was done on them. In this paper, we present the isolation and structures of eight compounds (**1–8**) from *Eria spicata* (D. Don) Hand.-Mazz. (*E. convallarioides*), which is distributed in Yunnan Province of China, Nepal, Sikkim, India, Burma, and Thailand [1]. The structures of the compounds were identified by a combination of spectroscopic methods (MS, ¹H, and ¹³C NMR) and comparisons with the literature data as *N*-methyl hordenine (**1**) [2], *N*-oxide hordenine (**2**), 3-(4-hydroxy-3-methoxyphenyl)-propenal (**3**) [3], nudol (**4**) [4], erianol (**5**) [4], 5 α ,8 α -epidioxy-(22*E*,24*R*)-ergosta-6,22-dien-3 β -ol (**6**) [5], β -sitosterol (**7**), and daucosterol (**8**). Previous investigation on this plant has yielded erianol, nudol, erianthridin, and β -sitosterol [4]. Compounds **1**, **2**, **3**, **6**, and **8** were isolated from this plant for the first time.

The whole plant of *E. spicata* was collected from Lianghe, Yunnan Province of China in February 2006, and was identified by Prof. Hong Yu (Yunnan University).

The dried whole plant of *E. spicata* (1.9 kg) was extracted with 70% MeOH (four times, each 15 L for 7 days) at room temperature. After concentration of the combined extracts, the resulting gummy material was suspended in water and then partitioned with EtOAc to afford EtOAc and aqueous residues. The EtOAc residue (35 g) was subjected to column chromatography over silicon gel and eluted with petroleum ether–EtOAc (10:1) to (0:1) to give seven fractions.

Fraction 2 (1.54 g) was subjected to column chromatography on silica gel eluted with petroleum ether–EtOAc (15:1) and then on a silica gel column developed with petroleum ether–CHCl₃ (10:1) repeatedly to obtain **5** (16 mg). Compound **7** (900 mg) was obtained from fraction 3 (1.58 g) by recrystallization. Fraction 4 (3.5g) was purified on a SiO₂ column eluted with petroleum ether–CHCl₃ (from 6:1 to 3:1), and each subfraction was purified on a SiO₂ column eluted with petroleum ether–CHCl₃ (4:1) repeatedly to obtain **3** (40 mg), **4** (230 mg), and **6** (18 mg). Fraction 7 (12.1 g) was separated on silica gel eluted with CHCl₃–MeOH (from 15:1 to 5:1) and then isolated on Sephadex LH-20 (MeOH–CHCl₃ 1:1) to afford **8** (60 mg).

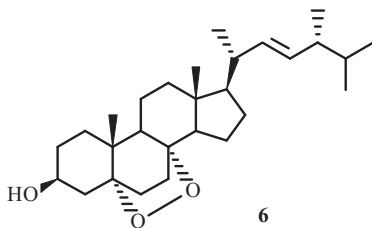
The aqueous residue was subjected to HPD-101 macroporous absorption resin eluted with H₂O, then with 95% ethanol. The H₂O portion (18.1g) was subjected to SiO₂ column chromatography eluted with CHCl₃–MeOH (from 5:1 to 0:1) to give four fractions. Fraction 2 (5.12 g) was purified on a SiO₂ column (CHCl₃–MeOH–H₂O 7:3:0.5) repeatedly to afford **2** (22 mg). Fraction 3 (3 g) was purified on SiO₂ column (CHCl₃–MeOH–H₂O 6.5:3.5:0.5) repeatedly to afford **1** (8 mg).

N-Methyl Hordenine (1). C₁₁H₁₈NO. ESI-MS 180 [M]⁺. ¹H NMR (500 MHz, CD₃OD, δ , ppm, J/Hz): 7.15 (d, J = 8.5, H-2', 6'), 6.74 (d, J = 8.5, H-3', 5'), 3.51 (t, J = 8.5, H-1), 3.02 (t, J = 8.5, H-2), 3.19 (9H, s, 3CH₃). ¹³C NMR (125MHz, CD₃OD, δ): 157.8 (s, C-4'), 131.0 (d, C-2', 6'), 127.3 (s, C-1'), 116.7 (d, C-3', 5'), 68.8 (t, C-1), 29.5 (t, C-2), 53.7 (q, 3CH₃).

N-Oxide Hordenine (2). C₁₀H₁₅NO₂. ESI-MS 181 [M]⁺. ¹H NMR (500 MHz, CD₃OD, δ , ppm, J/Hz): 7.16 (d, J = 8.5, H-2', 6'), 6.79 (d, J = 8.5, H-3', 5'), 3.37 (6H, s, 2CH₃), 3.34 (br.s, H-1), 3.02 (t, J = 8.5, H-2). ¹³C NMR(125 MHz, CD₃OD, δ): 157.3 (s, C-4'), 130.6 (d, C-2', 6'), 127.8 (s, C-1'), 116.5 (d, C-3', 5'), 59.9 (t, C-1), 43.4 (q, 2CH₃), 30.7 (t, C-2).

3-(4-Hydroxy-3-methoxyphenyl)-propenal (3). ¹H NMR (500 MHz, CDCl₃, δ , ppm, J/Hz): 9.56 (1H, d, J = 7.8, H-1), 6.53 (1H, dd, J = 15.8, 7.8, H-2), 7.32 (1H, d, J = 15.8, H-3), 7.19 (1H, s, H-2'), 7.03 (1H, d, J = 8.2, H-6'), 6.88 (1H, d, J = 8.2, H-5'), 3.84 (3H, s, 3'-OCH₃). ¹³C NMR (125 MHz, CDCl₃, δ): 153.0 (d, C-1), 114.9 (d, C-2), 193.5 (d, C-3), 126.6 (s, C-1'), 124.0 (d, C-2'), 146.9 (s, C-3'), 148.9 (s, C-4'), 109.4 (d, C-5'), 126.3 (d, C-6'), 55.9 (q, 3'-OCH₃). EI-MS *m/z* (%): 178 [M]⁺ (10), 149 (35), 85 (60), 71 (85), 57 (100).

1) Department of Chemistry, Yunnan Normal University, Street 121, Kunming 650092, P. R. China, e-mail: ygchen48@gmail.com; 2) Department of Natural Resources, Southwest Forestry University, Kunming 650224, Yunnan, P. R. China. Published in *Khimiya Prirodnikh Soedinenii*, No. 1, pp. 150–151, January–February, 2012. Original article submitted November 17, 2010.



Nudol (4). ^1H NMR (500 MHz, CDCl_3 , δ , ppm, J/Hz): 9.24 (1H, d, $J = 9.0$, H-5), 7.35 (2H, s, H-9, 10), 7.13 (1H, s, H-8), 7.12 (1H, d, $J = 9.0$, H-6), 7.03 (1H, s, H-1). ^{13}C NMR (125 MHz, CDCl_3 , δ): 109.2 (d, C-1), 148.0 (s, C-2), 141.9 (s, C-3), 151.5 (s, C-4), 129.7 (d, C-5), 116.9 (d, C-6), 154.5 (s, C-7), 112.0 (d, C-8), 127.3 (d, C-1), 126.6 (d, C-10), 133.9 (s, C-1'), 123.9 (s, C-2'), 118.9 (s, C-3'), 129.7 (s, C-4'), 60.1 (q, 3-OMe), 61.2 (4-OMe).

Erianol (5). ^{13}C NMR (125 MHz, CDCl_3 , δ): 36.9 (t, C-1), 30.9 (t, C-2), 76.2 (d, C-3), 40.3 (d, C-4), 46.7 (d, C-5), 26.6 (t, C-6), 117.5 (d, C-7), 139.1 (s, C-8), 49.6 (d, C-9), 34.8 (s, C-10), 21.4 (t, C-11), 39.3 (t, C-12), 43.3 (d, C-13), 54.9 (d, C-14), 22.9 (t, C-15), 27.9 (t, C-16), 55.9 (d, C-17), 11.8 (q, C-18), 14.1 (q, C-19), 36.3 (d, C-20), 18.9 (q, C-21), 30.4 (t, C-22), 37.0 (t, C-23), 38.7 (s, C-24), 152.4 (s, C-25), 109.3 (t, C-26), 19.4 (q, C-27), 15.1 (q, C-29), 27.5 (q, C-31), 27.2 (q, C-32).

5 α ,8 α -Epidioxy-(22E,24R)-ergosta-6,22-dien-3 β -ol (6). $\text{C}_{28}\text{H}_{44}\text{O}_3$. EI-MS: 428 $[\text{M}]^+$ (4), 410 $[\text{M} - \text{H}_2\text{O}]^+$ (5), 396 $[\text{M} - 32]^+$ (15), 251 (24), 152 (36), 81 (61), 69 (100). ^1H NMR (500 MHz, CDCl_3 , δ , ppm, J/Hz): 6.59 (d, $J = 8.3$, H-7), 6.50 (d, $J = 8.3$, H-8), 5.21 (d, $J = 7.7$, H-23), 5.16 (d, $J = 7.7$, H-24), 3.98 (m, H-3). ^{13}C NMR (125 MHz, CDCl_3 , δ): 34.6 (t, C-1), 30.0 (t, C-2), 66.4 (d, C-3), 36.9 (d, C-4), 82.1 (s, C-5), 135.4 (s, C-6), 130.7 (s, C-7), 79.7 (s, C-8), 51.1 (d, C-9), 36.9 (s, C-10), 23.4 (t, C-11), 39.3 (t, C-12), 44.5 (d, C-13), 51.6 (d, C-14), 19.9 (t, C-15), 28.6 (t, C-16), 56.3 (d, C-17), 12.8 (q, C-18), 18.1 (q, C-19), 39.7 (d, C-20), 20.8 (q, C-21), 135.2 (d, C-22), 132.3 (t, C-23), 42.7 (d, C-24), 20.6 (q, C-25), 33.0 (d, C-26), 19.6 (q, C-27), 17.5 (q, C-28).

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